

Bravo Metabolomics Workbench Off-site Plasma Metabolite Extraction Application Guide

For Research Use Only. Not for use in diagnostic procedures.



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About this guide

Overview

This guide describes the Off-site Plasma Metabolite Extraction application for the Bravo Metabolomics Sample Prep Platform. For more details on the Bravo Metabolomics Sample Prep Platform, see the *Getting Started Guide* in the Literature Library of the Bravo Metabolomics Workbench.

The procedures in this guide assume that you have been trained on how to operate the Bravo Platform.

WARNING

Using controls, making adjustments, or performing procedures other than those specified in the user documentation can expose you to moving-parts hazards and hazardous voltage. Before using the Bravo Platform, make sure you are aware of the potential hazards and understand how to avoid being exposed to them.

Software version

This guide documents the following versions or later:

- Bravo Metabolomics Workbench 1.0
- VWorks Automation Control 13.1.3
- Bravo Diagnostics 19.1

Related guides

Use this guide in conjunction with the following guides:

- Automation Solutions Products General Safety Guide. Provides general safety information and describes potential safety hazards that you might encounter when using Agilent Automation Solutions products. A copy of this safety guide is included with your shipment.
- *G5562A, G5563A Bravo Platform Safety and Installation Guide*. Describes potential safety hazards and how to avoid them, how to install the Bravo Platform, and how to install the Light Curtain and shields. A copy of this safety guide is included with your shipment.
- *Bravo Platform User Guide*. Explains how to set up, operate, and maintain the Bravo Platform and how to install accessories.

You can find the workbench user guides in the Literature Library of the Bravo Metabolomics Workbench software.

Contacting Agilent Technologies

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App description

The Off-site Plasma Metabolite Extraction application performs automated sample preparation on up to 96 plasma samples collected off site in a single protocol run. Before running this application, the plasma should have been premixed with a methanol/ethanol solution and transferred to one of the specified 96-well plate options. See "Labware requirements" on page 5. You can use the Reagent Transfer utility to prepare the labware.

During the Off-site Plasma Metabolite Extraction protocol, the Bravo Platform does the following:

- Lipid removal. Transfers the samples from a prepared Plasma with Methanol/ Ethanol plate to a Captiva EMR-Lipid plate, filters the samples to remove the lipids, and collects the eluate in a Collection plate.
- Eluate transfer to a final plate. Transfers a set volume of eluate from the wells in the Collection plate to a Final plate.

Next steps:

You dry-down the Final plate in a centrifugal evaporator, such as a Speedvac, for storage. Later, you can use the Reconstitution application to resuspend the sample.





*Assumes that the user manually pours the methanol and water for the Reconstitution app.

Before you start

This topic lists the required hardware, labware, and reagents for running the protocol.

Hardware requirements

The following figure and table show the primary hardware components for the Bravo Metabolomics Sample Prep Platform.

Figure Bravo Metabolomics Sample Prep Platform hardware components



| ltem | Name |
|------|---|
| 1 | Bravo Platform |
| 2 | Liquid-handling head, 96LT |
| 3 | Gripper upgrade |
| 4 | Heating Shaking Station (deck location 4) and STC controller |
| 5 | Filter Plate Holder (deck location 6) |
| 6 | Vacuum Filtration Station and Agilent ME4C NT VARIO Pump |
| | Manifold base (deck location 2) |
| | • Deep-well collar with black gasket on top (deck location 3) |
| 7 | Emergency-stop pendant |
| | Light Curtain and safety shields (not shown) |

Additional equipment

Centrifugal evaporator, such as a SpeedVac, to dry-down the eluate plate

Labware requirements



Using a labware type at a deck location other than an approved labware option can cause a collision resulting in equipment damage. Ensure that you use only an approved labware option for each deck location.

The following figure shows the nine Bravo deck locations for labware. The table lists the labware options that you can choose from for each deck location.

Figure Application labware locations on the Bravo deck (top view)

| l. EMR Collection Plate | 2. Vacuum Manifold (Base) | 3. Vacuum Manifold (Collar — Deep) |
|-----------------------------|---|---------------------------------------|
| ł. Empty | 5. Final Plate | 6. Captiva EMR-Lipio Plate |
| 7. Tip box (250 µL Tips) | 8. Plasma with Methanol/Ethanol Plate | 9. Tip box (250 µL Tips) |

 Table
 Labware options by deck location

| Deck location | Labware option | Manufacturer part number |
|------------------|---|-------------------------------|
| 1, 5, 8 | 96 Costar 3961 PP 2ml assay block | Corning Costar 3961 |
| 1, 5, 8 | 96 Agilent A696001000 Captiva collection plate | Agilent A696001000 |
| 5, 8 | 96 Agilent 203426-100 PP, 1 mL Rnd Btm | Agilent 203426-100 |
| 5 | 96 EK 2460 PP Rnd Well U Btm | E&K Scientific EK-2460 |
| 5 | 96 Greiner 655101 PS Clr Rnd Well Flat Btm | Greiner 655101 |
| 6 | 96 well Captiva EMR-Lipid Filter plate | Agilent 5190-1001 |
| | (Captiva EMR - Lipid plate) | |
| 7, 9 | 96 V11 LT Tip Box (250 µL disposable pipette tips) | Agilent 19477.002 |
| 8 | 96 Thermo Matrix 3741, V-bottom, 1 mL ScrewTop Storage Tubes (Plasma plate) | Thermo Fisher Scientific 3741 |
| | (| |

IMPORTANT

The protocol supports full plates or partial plates. Partial plates must be arranged in full, contiguous columns.

Samples and reagents

IMPORTANT

Prepared plasma premixed with methanol/ethanol

If required, use the Reagent Transfer utility to premix an internal standard in the Plasma with Methanol/Ethanol plate before the run.

Preparing the sample and reagent labware

To prevent evaporation, dispense the reagents into the labware immediately before running the protocol.

See "Labware requirements" on page 5. Ensure that the labware is prepared as follows:

- Fluids are arranged in full columns that are contiguous.
- Plasma columns start with column 1 of the tube rack or other labware. See "Labware requirements" on page 5 for deck location 8 options.
- Two tip boxes containing sufficient pipette tips for your run:
 - Tips for plasma transfer
 - Tips for eluate transfer

The columns of tips must start with column 1 in each tip box.

You can use the Reagent Transfer utility to transfer the fluids from one labware to another.

For partial-plate runs, you can use the Tip Transfer utility to arrange the pipette tips in the tip boxes.

See the following figure for an example of the labware layout for a partial-plate run.



Figure Example of labware layout for a run with four columns of plasma tubes

*You can specify the starting column of wells in the Captiva and Eluate plates.

Setting up the protocol

Opening the application

To open the application:

In the App & Utility Libraries page of the Bravo Metabolomics Workbench, locate Off-site Plasma Metabolite Extraction, and then click Open.

| C Works - [Workflow Library_v5.1.W/Form] | - 0 |
|---|-------------------------|
| Bravo Metabolomics Workbench: APP & UTILITY LIBRARIES | Agilent Trusted Answers |
| | App and Utility Library |
| Applications | |
| On-site Plasma Metabolite Extraction | Open |
| Off-site Plasma Metabolite Extraction | Open |
| Reconstitution | Open |
| Utilities | |
| Tip Transfer | Open |
| Reagent Transfer | Open |
| | |
| | |
| c - | |

Specifying the settings

Before starting the protocol, make sure the appropriate selections and values are specified in the Off-site Plasma Metabolite Extraction form.

| S VWe | rks - [Off-site | e Plasma Metabolite | Extraction_v5.2 | .VWForm] | | | | | | | | | | | |
|-------|-----------------|--------------------------------|------------------------|-----------|------------------------------|-----------------------|-----------------------|--------------|-------|---------------------------|--|---------------------------------------|----|-----------|------------------------|
| | Appl | lication | : Off-s | ite l | Plasma | a Me | tabolite | Extra | ction | | | 2 | | Agilent | Trusted Answers |
| | | Applicatio | on Settin | gs | | | | | | U N | Deck Layout | | | Status | Elapsed Time: 00:00:00 |
| | | Total Numbe Columns (1- | r of Sampl 12) 1 | e S Ci | tarting Wel aptiva EMR-Li | l Column pid Plate | (1-12) Final Plate | i | | 1. EMR Collec Plate | ction 2. Vacuum Manifold (Base) | 3. Vacuum Manifold (Collar – Deep) | Ì | 🗐 Instruc | tions |
| | | Settings | | | Step | 8 | | | | | | | | | |
| | | | | | 1. Lipid Re | moval | 2. Eluate 1 | ransfer | | 4. Empty | 5. Final Plate | 6. Captiva EMR-Lipid Plate | | Nun Ex | traction |
| | | Transfer Volu | me (2-1,000 |) | 500 | μί | 30 | p µL | | | | | | D Paus | se |
| | | Dispense Dist Well Bottom (| ance From (0.5-100) | | 2 | mm | 2 | mm | | 7. Tip box (250 µL Tip | 8. Plasma with Methanol/Ethano Plate | 9. Tip box (250 µL Tips) | | 🚚 Restore | e Defaults |
| | | Liquid Class | | 96 | disposable tip 5 | 1 - 200ul 💌 | 96 disposable tip | 51 - 200ul 💌 | | | | | | | |
| | | Filtration Time | D | | 180 | 580 | | | | | Labwara Tab | 0 | | 🛐 Toggle | Full Screen |
| | | Filtration Targ | jet Pressure | , | 200 | mbar | | | | Deck | | • | | 🔶 App and | Utility Library |
| | | Filtration Vent | t Delay | | 0 | sec | | | | Location | Labwar | а Туре | 7 | | |
| | | Filtration Time | e | | 60 | 600 | | | | 1 | 96 Agilent A696001000 Captiva | collection plate | · | | |
| | | (blowout) | | | 1 | 300 | | | | 2 | Vacuum Manifold (Base) | | | | |
| | | Filtration Targ (blowout) | jet Pressure | , | 400 | mbar | | | | 3 | Vacuum Manifold (Collar | - Deep) | | | |
| | | Filtration Vent | t Delay | | 15 | sec | | | | 4 | Empty | | - | | |
| | | (blowout) | | | 1 | 500 | | | | 5 | 96 Agilent 203426-100 PP, 1 m | Rnd Btm | ·] | | |
| | | | | | | | | | | 0 | Captiva ElviR-Lipid Flate | - 10477 002 | | | |
| | | | | | | | | | | , | 05 Aniest 202425-100 BB 1 m | n 194/7.002) | | | |
| | | | | | | | | | | 0 | 96 V11 LT 260 Tin Part (a | (= 10477 002) | 1 | | |
| | | | | | | | | | | | 50 VII LI 250 IIP Box (p. | 1 194/7.002) | | | |

To specify the settings:

- 1 *Optional.* Click **Restore Defaults** to set all the form settings to their defaults.
- 2 Under Application Settings, specify the following:

| Setting | Description | | | | |
|---|--|--|--|--|--|
| Total Number of Sample Columns (1–12) | The number of columns of plasma samples in the tube rack at deck location 8, for example, type 12 for a full rack of samples. | | | | |
| (1 12) | IMPORTANT Ensure that the tubes are in full columns that are contiguous, starting with column 1. | | | | |
| | Default: 1 | | | | |
| Starting well column | n (1–12), Default: 1 | | | | |
| Captiva EMR-Lipid Plate | The starting column of the destination wells in the Captiva plate on the Vacuum Filtration Station at deck location 2. For a full-plate run, this is column 1. | | | | |
| | The protocol transfers the sample from the Plasma with Methanol/Ethanol plate to the Captiva plate, starting at this column number. | | | | |

Setting up the protocol

| Setting | Description |
|-------------|---|
| Final Plate | The starting column of wells in the plate at deck location 5. For a full-plate run, this is column 1. |
| | The protocol transfers a set volume of eluate from the EMR Collection plate to the Final plate, starting at this column number. |

- **3** Specify the **Settings** for the following **Steps**:
 - 1. Lipid Removal
 - 2. Eluate Transfer

1. Lipid Removal

| Step | Description |
|---------------------------------------|--|
| Transfer Volume | The volume of liquid per pipette tip to transfer from the Plasma with Methanol/Ethanol Plate (deck location 8) to the Captiva EMR-Lipid plate (deck location 2). Default: 500 (µL) Range: 2–1000 (µL) |
| Dispense Distance From Well Bottom | The dispense distance between the end of the pipette tips and the well bottoms during the liquid transfer. Default: 2 (mm) Range: 0.5–100 (mm) |
| Liquid Class | The pipetting speed and accuracy for the liquid transfer. You may choose from the options, which are based on the tip type and volume being transferred. These are good general-purpose liquid classes for most reagents: 96 disposable tip 1 -2 μL 96 disposable tip 2 - 50 μL 96 disposable tip 51 - 200 μL |
| Filtration Time | The length of time, in seconds, to leave the vacuum on. At the end of the period, the vacuum will turn off. Default: 180 (s) Range: 0–86400 (s) (24-hour maximum) |
| Filtration Target Pressure | The difference between the pressure of the outside atmosphere above the filter and the pressure in the Vacuum Filtration Station manifold, including the enclosure beneath the filter. For example, if you set the Target pressure to 600 mbar and the ambient pressure displayed on the VARIO pump is 1000 mbar, the vacuum will remain on until the reading on the VARIO pump reaches 400 mbar. Default: 200 (mbar) Range: 0–10000 (mbar) |

| Step | Description |
|------------------------------------|---|
| Filtration Vent Delay | The length of time, in seconds, to wait for the air pressure under the filter to equalize with the ambient air pressure. |
| | Default: 0 (s) |
| | Range: 0–86400 (s) (24-hour maximum) |
| Filtration Blowout | A second filtration task to ensure that all the residual fluid has passed through the Captiva plate (blowout) on the Vacuum Filtration Station. |
| Filtration Time | The length of time to leave the vacuum on. |
| (blowout) | Default: 60 (s) |
| | Range: 0–86400 (s) (24-hour maximum) |
| Filtration Target Pressure | The pressure (mbar) to use to filter any remaining residual fluid through the Captiva plate. Typically, this would be a higher value that used for the primary filtration task. |
| (blowout) | Default: 400 (mbar) |
| | Range: 0–10000 (mbar) |
| Filtration Vent Delay (blowout) | The length of time (seconds) to wait for the air pressure under the filter to equalize before the Bravo gripper attempts to disassembly the Vacuum Filtration Station. |
| | Default: 15 (s) |
| | Range: 0–86400 (s) (24-hour maximum) |

2. Eluate Transfer

| Step | Description |
|---------------------------------------|--|
| Transfer Volume | The volume of liquid per pipette tip to transfer from the EMR Collection plate (deck location 1) to the Final plate (deck location 5). |
| | Default: 300 (µL) |
| | Range: 2–1000 (µL) |
| Dispense Distance From Well Bottom | The dispense distance between the end of the pipette tips and the well bottoms during the liquid transfer. Default: 2 (mm) Range: 0.5–100 (mm) |
| Liquid Class | The pipetting speed and accuracy for the liquid transfer. You may choose from the options, which are based on the tip type and volume being transferred. These are good general-purpose liquid classes for most reagents: 96 disposable tip 1 -2 μL 96 disposable tip 2 - 50 μL 96 disposable tip 51 - 200 μL |

Specifying the labware



Using a labware type at a deck location other than an approved labware option can cause a collision resulting in equipment damage. Ensure that you use only an approved labware option for each deck location.

To specify the labware:

1 In the form, refer to the **Deck Layout**.

| EMR Collection Plate | 2. Vacuum Manifold (Base) | 3 Vacuum Manifold (Collar – Deep) |
|-----------------------------|---|--------------------------------------|
| 4. Empty | 5. Final Plate | 6. Captiva EMR-Lipid Plate |
| 7. Tip box (250 µL Tips) | 8. Plasma with Methanol/Ethanol Plate | 9. Tip box (250 µL Tips) |

2 In the Labware Table, select the labware you are using for deck locations 1, 5, and 8.

| Deck Location | Labware Type | |
|------------------|--|---|
| 1 | 96 Agilent A696001000 Captiva collection plate | • |
| 2 | Vacuum Manifold (Base) | |
| 3 | Vacuum Manifold (Collar - Deep) | |
| 4 | Empty | |
| 5 | 96 Agilent 203426-100 PP, 1 mL Rnd Btm | • |
| 6 | Captiva EMR-Lipid Plate | |
| 7 | 96 V11 LT 250 Tip Box (p/n 19477.002) | |
| 8 | 96 Agilent 203426-100 PP, 1 mL Rnd Btm | • |
| 9 | 96 V11 LT 250 Tip Box (p∕n 19477.002) | |

Running the protocol

Before you start

- Prepare the samples and reagents. See "Preparing the sample and reagent labware" on page 6.
- Ensure that you have two boxes of fresh pipette tips containing the required tips for your run:
 - Full-plate run. Use two full tip boxes.
 - **Partial-plate run**. Two tip boxes with the corresponding number of pipette tips arranged in contiguous columns starting at column 1.

About performing a mock run (optional)

If you are unfamiliar with the protocol and would like to see how it operates and troubleshoot problems before running it with valuable samples and reagents, you can perform a mock run using empty labware.

You prepare for a mock run the same way you would prepare for a real protocol run, except that you use empty labware for a totally dry run or labware containing water for a wet run. To decrease the run time, you can decrease the volumes and filtration time.

Starting the protocol run

CAUTION

To start the protocol run:

- 1 Review the selections in the protocol form to confirm they are correct.
- 2 Verify that the physical layout on the Bravo deck matches the **Deck Layout** image in the form. Make sure the labware are properly seated within the platepads on the Bravo deck.

Improperly seated labware can cause a hardware collision, resulting in equipment damage. Ensure that all labware are properly seated within the alignment features of their respective platepads.

- 3 Click Nun Extraction
 - If this is the first time the utility has been run after powering up the Bravo Platform, the device initialization process begins. Proceed to step 4.
 - If the platform is already initialized, skip to step 6.

4 If the **Bravo Error** message appears stating **There appears to be a plate present**, verify that Bravo gripper is not holding labware, and then

click Ignore and Continue, leaving device in current state to continue the initialization.

| the gripper's plate presence senso Choose "Retry" to check the pla again. | nt in, or in front of ir. ate presence sensor | ^ |
|--|---|----|
| Choose "Ignore" to continue to there is a plate currently held by t remove it first. It will be dropped it | nome the Gaxis. If he gripper, please f G homing is | |
| successful, and may get stuck if n - Choose "Abort" to cancel initial: | ot. zation. | |
| | | 5 |
| | | Y |
| | | |
| Diagnostics | | |
| <u>Retry</u> | | _ |
| <u>Diagnostics</u> | levice in current state | -, |
| <u>Retry</u> <u>Ignore and Continue, leaving of Abort</u> | levice in current state | |

5 If the **Please verify that it is safe to home the W-axis** message appears, click **Retry** to continue homing the pipetting axis (*w*-axis).

| Please verify that it is safe to hom aspirate/dispense axis). If there is may want to manually home the W over a waste position. - Choose "Retry" to continue hom - Choose "Ignore" to leave the W - Choose "Abort" to cancel initials | e the W-axis (the s fluid in the tips you -axis in diagnostics ang the W-axis. -axis unhomed. zation. | < | 1 |
|--|--|---|---|
| Dagnostics | | Ý | |
| Retry Ignore and Continue, leaving o | levice in current state | | |
| | Add to Error Librar | y | |

When the initialization process is finished, the orange lights on the Bravo Platform light panel flicker briefly and then begin to flash.

- **6** When the **Bravo Deck Setup** dialog box appears, verify that the deck layout is correct.
 - If it is correct, click Continue.
 - If it is not correct, click **Pause and Diagnose**, and then click **Abort process** in the Scheduler Paused dialog box. Resolve the problem, and then restart the run.

| Position 1: EMR Collect Position 2: Vacuum Fil Position 3: Vacuum Fil Position 4: Empty Position 5: Final Plate Position 5: Captiva EN Position 7: Tips for Pla Position 9: Plasma with | tion Plate tration Base tration Collar (Deep) 4R-Lipid Plate asma b Methanol /Ethanol |
|--|--|
| Position of Plasma wit | n Methanol/Ethanol |
| Plate Position 9: Tips for Fir | nal Plate. |

- 7 If an Error message appears, the software detected a conflict in the protocol setup.
 - **a** Follow the on-screen instructions to click **Pause and Diagnose** and then click **Abort process** in the Scheduler Paused dialog box.
 - **b** Resolve the conflict described in the error message, for example:

| ERROR | |
|--|--|
| ERROR: The Total Nu Columns must have a available well columns Captiva EMR-Lipid an | mber of Sample n equal number of s to process the d Final Plates. |
| Please dick on 'Pause and Diagnose' then 'Abort Process'. | |
| | |
| | 1 |
| Pause and Diagnose | Continue |

- Insufficient number of columns of pipette tips
- Unequal number of columns selected for source and destination plates



To monitor the progress of the run, check the **Status** area of the form.

| Status | Elapsed Time: 00:05:16 |
|--------------------|------------------------|
| Protocol Complete! | |
| | |
| | |

About stopping or pausing a run

IMPORTANT

Attempting to pause a running protocol to change a setting can be detrimental to the protocol. If you need to change a setting in a protocol that is actively running, pause the protocol, select Abort process from the Scheduler Paused dialog box, change the setting, and then restart the protocol.

For more detailed instruction, see the *Bravo Metabolomics Sample Prep Platform Getting Started Guide*.

Cleaning up

When the protocol run is finished:

- Remove all labware from the Bravo deck.
- Discard solutions, organic waste, and used labware following appropriate waste disposal procedures.

WARNING

Make sure you discard the chemical waste and used labware according to your lab's waste disposal procedures and in compliance with all local, state, and federal safety regulations.

Automation movements during the protocol

This section describes the basic movements of the Bravo Platform during the protocol using the default protocol settings. Changing the selections or parameters will alter the movements.

| Protocol step | Head moves to deck location | Action |
|--|-----------------------------------|---|
| Vacuum Filtration Station Assembly | 1, 2 | Moves the EMR Collection plate to the manifold base at deck location 2. |
| | 3, 2 | Moves the Deep Well Collar to the manifold base at deck location 2. |
| | 6, 2 | Moves the Captiva plate to the Vacuum Filtration Station at deck location 2. |
| Tip Pick Up | 7 | Presses on the columns of pipette tips from the tip box. The number of full columns corresponds to the specified number of columns in the Plasma plate. |
| Plasma Transfer | 8, 2 | Transfers the sample from the Plasma with Methanol/Ethanol plate into the specified columns of wells in the Captiva plate. |
| | | <i>Note</i> : The transfer cycle may repeat depending on the volume to be transferred. |
| Filtering | 2 | Filters the sample through the Captiva plate into the EMR Collection plate at the Vacuum Filtration Station. |
| Tip Removal | 7 | Ejects the used pipette tips into the tip box on deck location 7. |
| Vacuum Filtration Station Disassembly | 2, 6 | Moves the Captiva plate to deck location 6. |
| | 2, 3 | Moves the Deep Well Collar to deck location 3. |
| | 2, 1 | Moves the EMR Collection plate to deck location 1. |
| Tip Pick Up | 9 | Presses on the columns of pipette tips from the tip box. The number of full columns corresponds to the specified number of columns in the Final plate. |
| Sample Transfer | 1, 5 | Transfers a specified volume of sample from the EMR Collection plate into the specified wells of the Final plate. |
| | | <i>Note</i> : The transfer cycle may repeat depending on the volume to be transferred. |
| Tip Removal | 9 | Ejects the used pipette tips into the tip box at deck location 9. |

Bravo Metabolomics Workbench

Automation movements during the protocol