## **Agilent QuEChERS Kits**

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**Product Manager SPE** 

**EPRW 2010** 

## What is QuEChERS (pronounced "Catchers")

- Quick, Easy, Cheap, Effective, Robust and Safe
- Sample Preparation method developed jointly by US FDA and EU Food Regulatory Agencies
- Methodology for simplified extraction of a large number of pesticide residues in fruits and vegetables
- QuEChERS is a fast sample preparation method for screening for analytes of interest, not for concentrating samples
- QuEChERS is still in it's "Infancy"; being adopted worldwide

	Luke method or traditional SPE	QuEChERS	Bottom Line
Estimated Time to process 6 samples (min)	120	30	4x faster
Solvent Used (mL)	90 mL	10mL	9 x less solvent
Chlorinated Waste (mL)	30 mL	none	safer, greener, less costly
Glassware/ specialized equipment	Funnel, water bath, 200mL containers, evaporator, etc.	None	No additional supplies needed

QuEChERS Brochure: literature #5990-3562EN

## Comparison of the 3 Methods

- Original QuEChERS method (unbuffered)
  - 4 or 6 g MgSO4, 1 or 1.5 g NaCl
- AOAC method 2007.01 (AOAC)
  - 6 g MgSO4, 1.5 g Na Acetate
- EN method 15662 (CEN)
  - 4 g MgSO4, 1 g NaCl, 1 g NaCitrate, 0.5 g disodium citrate sesquihydrate

All 3 methods give excellent results: average 98% recoveries with 10% RSDs

Ionic Strength of the two methods:

- AOAC buffered method
  - Relatively strong acetate buffering conditions (pH 4.8)
- EN buffered method
  - Weaker citrate buffering conditions (pH 5-5.5)

#### **Universal d-SPE**

- 50 mg PSA, 50 mg C18, 7.5 mg GCB, 150 mg MgSO4
- Cleanest extract, for all matrices without unacceptably affecting recoveries even for structurally planar pesticides
- J. Chromatography A 1217 (2010) 2548-2560.

#### **Extraction Choice**

AOAC over EN over Unbuffered

## The Value of Agilent SampliQ QuEChERS Kits

- Reduced time and labor: New Ceramic Homogenizers
  - · With addition of ceramic homogenizers in tubes, shaking step is minimized to only 20 seconds
  - Saves you time, money and improves results
  - Proof of performance included in QuEChERS brochure (5990-3562EN)

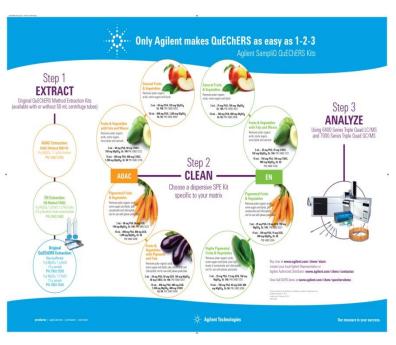


- Anhydrous foil packs ensure salt integrity, under nitrogen
- Avoid exothermic reactions by adding salts after solvent
- Exothermic reactions cause samples to degrade
- No need for additional centrifuge tubes



- Select an extraction kit based on methodology (AOAC or EN)
- Select a dispersive kit based on food type (meat, fruits, vegetables, etc)
- Extraction packets available without tubes
- Technical support: Over 20 application notes on QuEChERS:
  - All available on our website: <u>www.agilent.com/sampliQ</u>
  - QuEChERS Food Application "Notebooks": Lit number: 5990-4977EN
  - Seminars and hands-on training available worldwide





Backed by the Agilent brand you know and trust for all your instrument and column needs!



#### SampliQ QuEChERS Market Leader

- ✓ Total Extraction Kit Options: 16
- ✓ Total Dispersive Kit Options: 33
- ✓ Applications: 20+
- ✓ Local support (tech support specialists, product specialists and distributors throughout Europe)
- √ Solutions expertise (GC/MS and LC/MS)
- √ Certificate of Composition with each kit
- ✓ Satisfied customers around the world!

The most complete QuEChERS solution for customers in the market today

#### **Extraction Kit Packaging**

- ✓ Water and UV resistant packaging 3 layer material
- ✓ Pack the salts under nitrogen not vacuum
- ✓ Increases pouring capabilities
- √ Substantially reduces clumping of salts
- ✓ All ingredients listed on packet

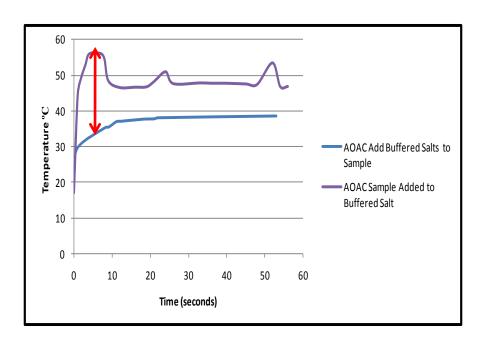
Packaging salts separately allows customers to add sample before adding salts

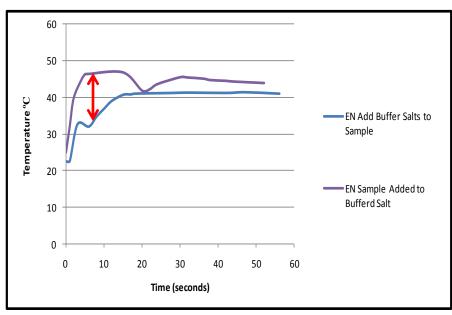
Prevents exothermic reaction Prevents degradation of sample Ensures maximum recoveries





# Temperature Graphs from AOAC and EN Extraction Kits





Represent the thermal data acquired from the AOAC and EN extraction kits. A substantial increase in temperature is observed (increase of approx 25 °C for the AOAC method and approx 10 °C for the EN method) when the produce sample is added to the extraction salt in the 50 mL PP tube versus when the extraction salt from an anhydrous pouch is added to the produce sample

#### **Effect of Temperature on Recovery**

The following four pesticides were used to evaluate the thermal effects that are prevalent when the produce is added to the extraction salts

H<sub>3</sub>C CH<sub>5</sub>



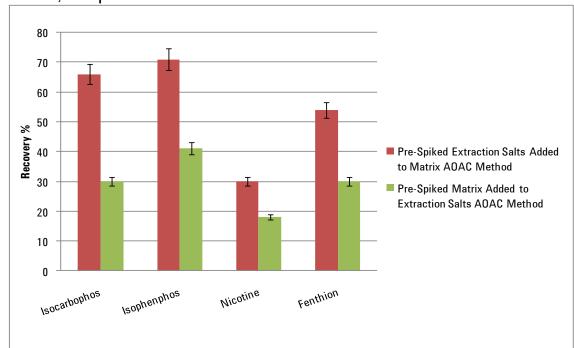
Isocarbophos

Isofenphos

**Fenthion** 

Nicotine\*

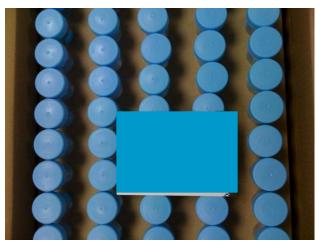
Analysis was performed on a Agilent 6410 Triple Quad LC/MS using a Phenyl- Hexyl column 2.1 x 50 mm, 3.5 µm



Represents the negative effect adding the produce matrix to the salt has on recovery during the QuEChERS extraction step, AOAC method

## Other Companies' Extraction Packaging







#### Improvements to the Extraction of Samples

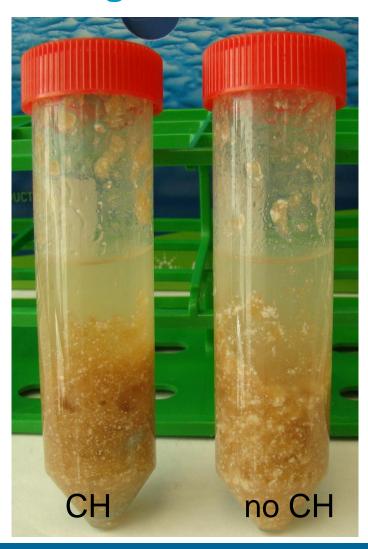
#### **Step 1:** Extraction salts with comminuted fruit/vegetable

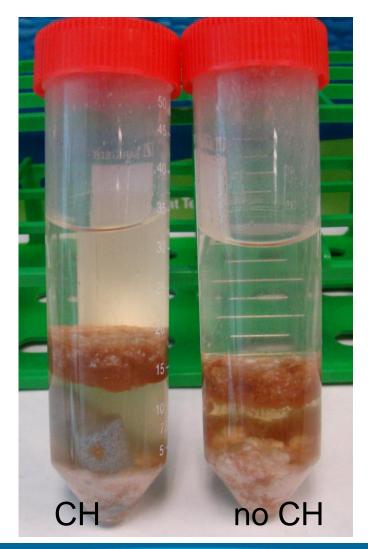
- Consistency in shaking, everyone shakes differently
- Variability in QuEChERS applications, recovery and RSDs

- SampliQ Ceramic Homogenizers
  - Reduces shaking time from 1 minute to <20 seconds!</li>
  - Consistent extraction of the sample with the salts
  - Breaks up salt agglomerates
  - Facilitates homogenization with angle cut
  - Increase recovery of pesticides from sample
  - 3 different sizes: 50 mL extraction tube and 2 dispersive tubes 15 and 2 mL



# **QuEChERS Extraction: With and Without Ceramic Homogenizers**

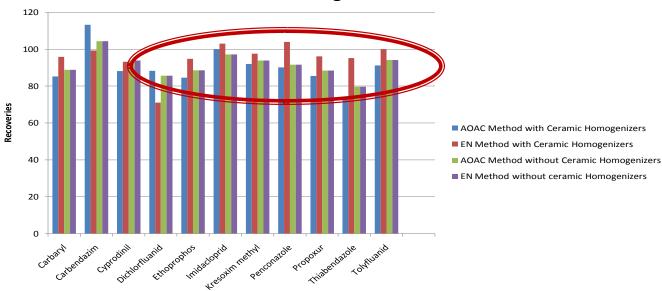


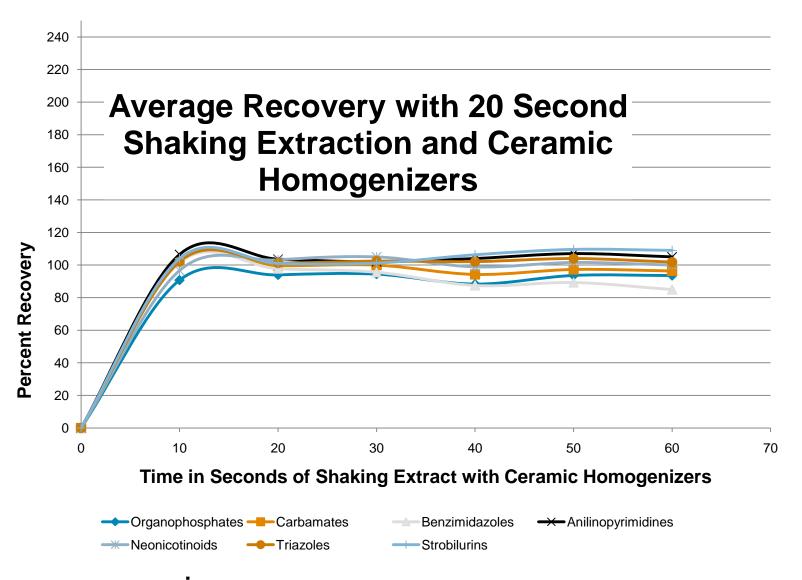


#### **Ceramic Homogenizers**

- Inert ceramic material
- No loss in pesticide recovery
- Consistent recovery

#### Comparison of QuEChERS Method AOAC and EN With and Without Ceramic Homogenizers





Pesticides used in study Acephate, Carbaryl, Carbendazim, Cyprodinil, Imidacloprid, Imazalil, Methamidophos, Penconazole, Propoxur, Pymetrozine, Thiabendazole, Thiophanate-methyl, Ethoprophos, Kresoxim-methyl; Apple matrix

# **QuEChERS applications in 2010: beyond just Pesticides in Fruits and Vegetables**

Application	Literature	QuEChERS Kit Used
Veterinary Drugs (antibiotics) in animal tissues	5990-5085EN 5990-5086EN	5982-5650 5982-4921
PAHs in Fish	5990-5411EN	5982-5755 5982-5058
Pesticides in Olive Oil	5990-5553EN	5982-5550 5982-5122
PAH's in Soil	5990-5452EN	5982-5755 5982-5156
Acrylamides in Fried Food	5900-5940EN	5982-5850 5982-5022
Pesticides in Baby Food	5990-5028EN	5982-5755 5982-5158
PCBs in Fish and Fish Oil Supplements	Coming soon!	
Pesticides in Essential Oils	Coming soon!	
Drugs in Biologicals	Coming Soon!	



Learn more today at www.agilent.com/chem/SampliQ

## **Agilent's' Solutions to Customers' Concerns**

Concerns	Agilent's Solution	Agilent Advantage
Salts in a tube	Salts in extraction packet	
Extra 50 mL tube	Empty 50 mL tubes	
Storage and stock of extraction and dispersive tubes	Organized box arrangement and extraction packets	
Shaking variability	Ceramic homogenizers	
Making standards: time and consistency	Standards available	

## **Appendix**



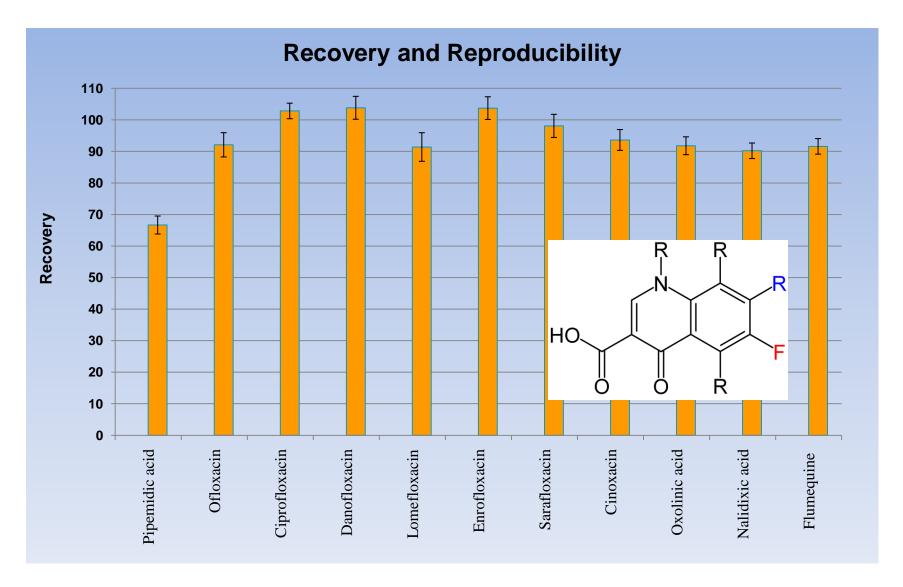
## Agilent SampliQ's QuEChERS Research

- Expanding QuEChERS Methodology!
  - ✓ Pharmaceutical
    - Veterinary Drugs in Animal Tissue
  - Environmental and Advancements in Food
    - Pesticides in Olive Oil
    - Pesticides in Coffee
    - PAHs in Fish and Soil
    - Acrylamides in Fried Foods

# **Cutting Edge Applications: Veterinary Drugs in Animal Tissue**

# Determination of Quinolone and Sulfonamide antibiotics in bovine liver:

- Extraction was performed using SampliQ EN extraction kit
- SampliQ dispersive SPE kits (25 mg C18\* and 150 mg MgSO<sub>4</sub>), for drug residue in animal tissue (NEW)
- Analyzed by LC/MS/MS



Recoveries for the 11 Quinolones from Bovine Liver



## **QuEChERS Extraction Procedure: Antibiotics in**

Liver

Weigh 2 g homogenized liver sample (± 0.05 g) in 50 mL centrifuge tube.

Spike 50 µL of IS spike solution, 50 µL of QC spike solution if necessary. Vortex 30 s.

Add 8 mL of water. Vortex.

Add 10 mL of 1% AA in ACN, and shake vigorously for 30 s.

Add SampliQ EN QuEChERS extraction kit and shake vigorously for 1 min.

Centrifuge @ 4000 rpm for 5 min.

Transfer 6 mL of upper ACN layer to SampliQ EN QuEChERS fatty dispersive-SPE 15 mL tube.

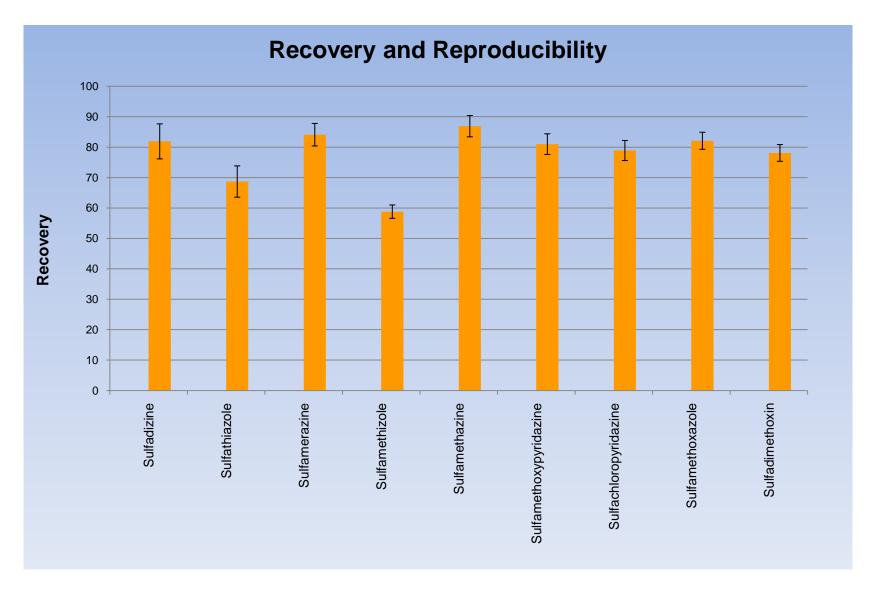
Vortex 2 min. Centrifuge @ 4000 rpm for 5 min.

Transfer 4 mL extract to another tube. Blow down @ 40 °C with  $N_2$  .

Reconstitute into 800 µL 1:9 MeOH/H<sub>2</sub>O with 0.1% FA. Vortex and sonicate.

Filter samples with 0.22 µm cellulose acetate spin filter.

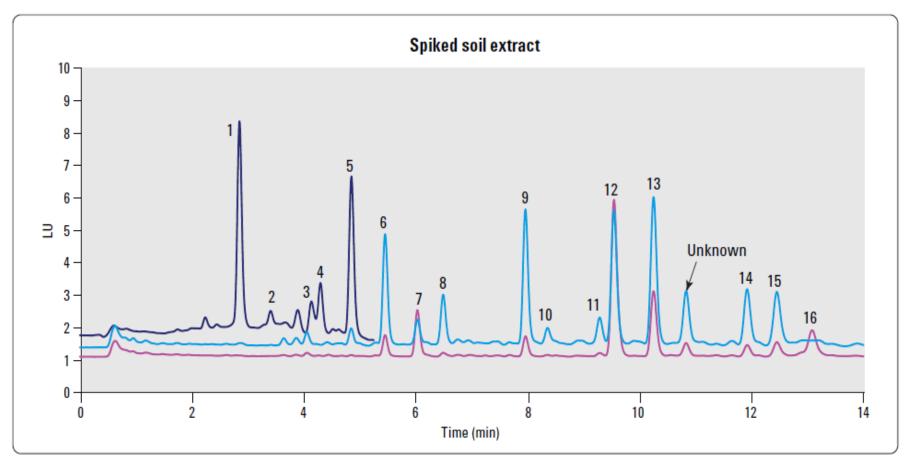
Samples are ready for LC/MS/MS analysis.



Recoveries for the 9 Sulfonamides in Bovine Liver



# PAHs in Soil Employing QuEChERS Extraction and HPLC-FLD Analysis



Overlay HPLC — FLD chromatograms of the spiked soil sample containing: 1. Nap 2. Acy 3. Ace 4. Flu 5. Phe 6. Ant 7. Fln 8. Pyr 9. BaA 10. Chr 11. BeP 12. BeA 13. BkF 14. DahA 15. BghiP 16. InP. The spiking level for this sample was a level 1 (see Table 3). The blue portion of the chromatogram used the following excitation/emission wavelengths: 260-nm/352-nm; the red portion: 260-nm/420-nm and the light blue portion: 260-nm/440-nm.



# PAHs in Soil Employing QuEChERS Extraction and HPLC-FLD Analysis: Recovery and RSDs

РАН	Spiking level (ng/g)		
	1	2	3
Naphthalene	20	100	200
*Acenaphthylene	20	100	200
Acenaphthene	10	50	100
Fluorene	10	50	100
Phenanthrene	10	50	100
Anthracene	10	50	100
Fluoranthene	10	50	100
Pyrene	10	50	100
1,2-Benzanthracene	5	20	50
Chrysene	10	50	100
Benzo[e]pyrene	5	20	50
Benz[e]acenaphthylene	5	20	50
Benzo[k]fluoranthene	5	20	50
Dibenzo[a,h]anthracene	5	20	50
Benzo[g,h,i]perylene	5	20	50
Indeno[1,2,3-cd]pyrene	5	20	50

<sup>\*</sup> UV detection at 230 nm

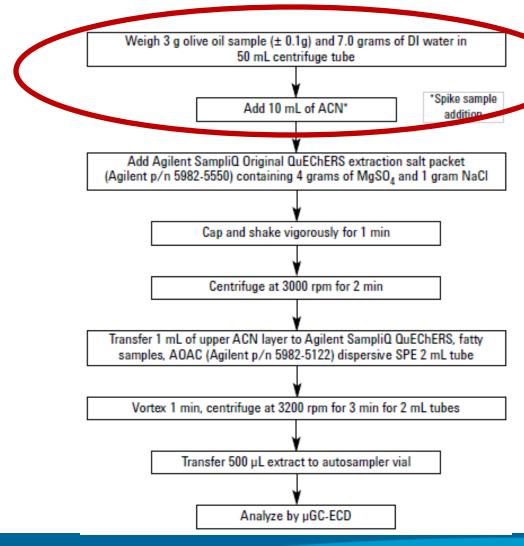
			Level of spiking	(ng/g) $(n = 6)$			
	1		2		3	3	
	%Recovery	%RSD	%Recovery	%RSD	%Recovery	%RSD	
Naphthalene	96.5	0.7	86.2	1.4	92.8	1.4	
*Acenaphthylene	87.3	0.7	90.0	1.3	91.7	1.6	
Acenaphthene	91.0	1.8	89.2	1.1	89.7	1.4	
Fluorene	95.2	0.8	91.4	1.3	86.0	1.2	
Phenanthrene	93.0	1.0	94.6	0.7	98.1	0.9	
Anthracene	91.9	1.1	90.0	0.8	97.6	0.7	
Fluoranthene	93.5	1.7	94.7	1.3	87.9	1.5	
Pyrene	96.3	1.3	89.4	0.9	91.2	1.9	
1,2-Benzanthracene	92.9	1.7	87.8	1.5	92.8	0.7	
Chrysene	98.0	1.4	92.4	1.2	95.8	1.0	
Benzo[e]pyrene	97.2	1.0	97.5	0.7	90.3	8.0	
Benz[e]acenaphthylene	93.2	0.9	93.1	0.6	98.0	0.7	
Benzo[k]fluoranthene	94.1	1.1	97.6	0.7	91.4	1.1	
Dibenzo[a,h]anthracene	89.2	1.0	99.2	1.7	90.8	1.3	
Benzo[g,h,i]perylene	91.0	0.9	96.7	0.8	97.3	1.6	
Indeno[1,2,3-cd]pyrene	86.0	1.2	97.8	0.8	94.3	1.3	

<sup>\*</sup> UV detection at 230 nm

PAHs in Soil using QuEChERS extraction and Agilent 5975T GC-MS

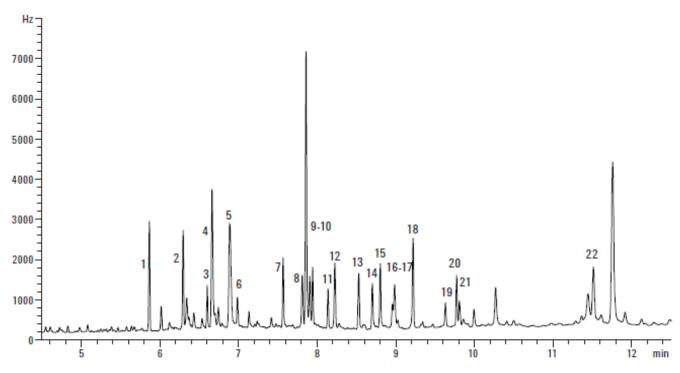


# Pesticides in Olive Oil by Dual Column GC-µECD Analysis: QuEChERS Procedure



## Dual Column GC-µECD Primary Column Analysis

Agilent J&W DB-35ms: 80 ng/mL CLP pesticide spiked olive oil



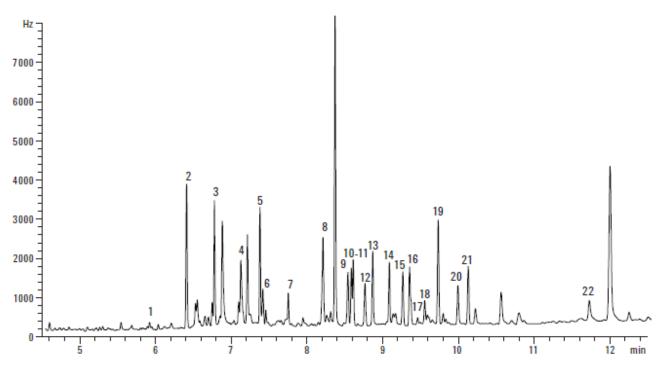
- Tetrachloro-m-xylene (surrogate standard)
- α-BHC
- y-BHC
- β-BHC
- Heptachlor
- $\delta$ -BHC
- Aldrin
- 8. Heptachlor epoxide
- 9. y-chlordane
- 10. α-chlordane
- 11. Endosulfan I

- 12. Dieldrin
- p,p-DDE
- 14. Endrin
- 15. p,p-DDD
- 16. Endosulfan II
- p,p-DDT
- 18. Endrin aldehyde
- Endosulfan sulfate
- 20. Methoxychlor
- 21. Endrin ketone
- 22. Decachlorobiphenyl (surrogate standard)



## GC-µECD Second Column: Confirmation Analysis





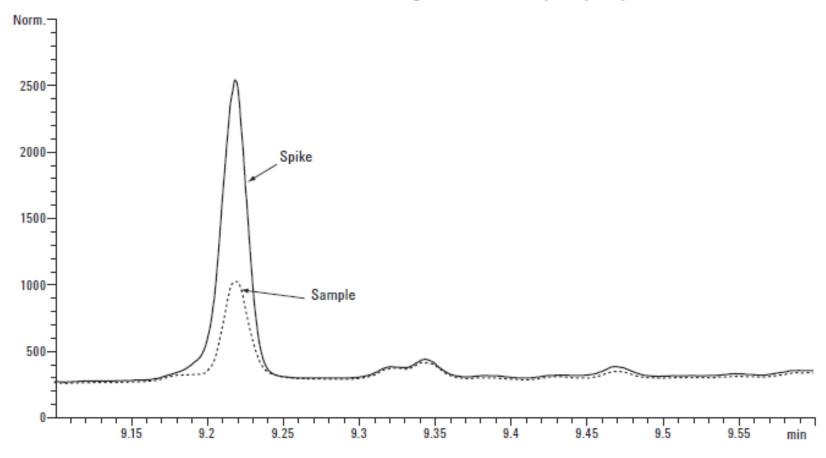
- Tetrachloro-m-xylene (surrogate standard)
- α-BHC
- y-BHC
- B-BHC
- Heptachlor
- δ-BHC
- Aldrin
- 8. Heptachlor epoxide
- 9. y-chlordane 10. α-chlordane
- 11. Endosulfan I

- p,p-DDE
- 13. Dieldrin
- 14. Endrin
- 15. p,p-DDD
- 16. Endosulfan II
- 17. Endrin aldehyde
- 18. p,p-DDT
- Endosulfan sulfate
- 20. Methoxychlor
- 21. Endrin ketone
- 22. Decachlorobiphenyl (surrogate standard)

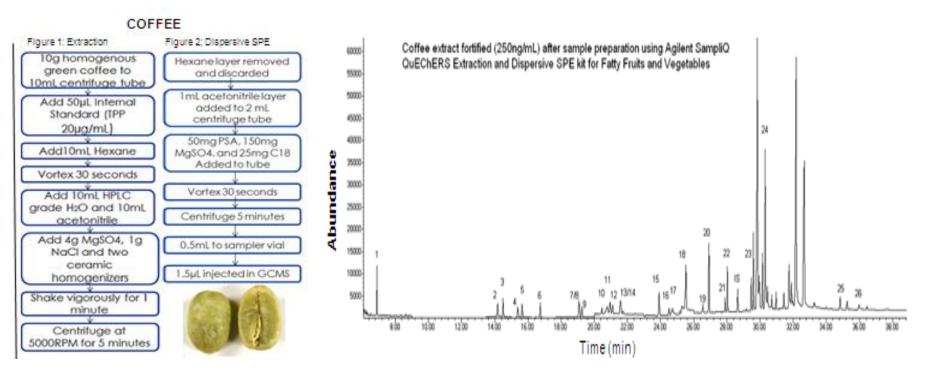


## GC-µECD Identification of Pesticide in Olive Oil

#### Endosulfan sulfate identified in olive oil on the Agilent J&W DB-35ms primary analysis column



#### **QuEChERS Extraction for Pesticides in Coffee**



Dr. Karyn Usher, West Chester University



# PAHs in Fish Employing QuEChERS Extraction and HPLC-DAD Analysis: Recovery and RSDs

PAH	Spiking level (ng/g)		
	1	2	3
Naphthalene	20	100	200
*Acenaphthylene	20	100	200
Acenaphthene	10	50	100
Fluorene	10	50	100
Phenanthrene	10	50	100
Anthracene	10	50	100
Fluoranthene	10	50	100
Pyrene	10	50	100
1,2-Benzanthracene	5	20	50
Chrysene	10	50	100
Benzo[e]pyrene	5	20	50
Benz[e]acenaphthylene	5	20	50
Benzo[k]fluoranthene	5	20	50
Dibenzo[a,h]anthracene	5	20	50
Benzo[g,h,i]perylene	5	20	50
Indeno[1,2,3-cd]pyrene	5	20	50

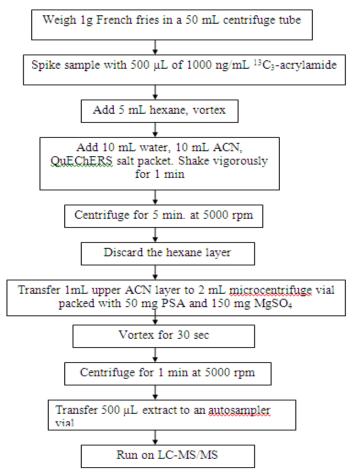
<sup>\*</sup> UV detection at 230 nm

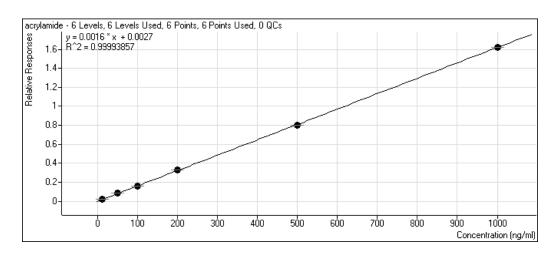
PAH			Level of spiking	(ng/g) (n = 6)		
	1		2		3	
	%Recovery	%RSD	%Recovery	%RSD	%Recovery	%RSD
Naphthalene	94.7	1.4	97.9	1.1	93.8	1.4
*Acenaphthylene	87.8	1.7	96.3	1.2	85.6	0.8
Acenaphthene	92.1	1.5	93.0	1.8	96.7	0.8
Fluorene	98.1	1.5	89.9	1.0	97.2	0.9
Phenanthrene	90.6	0.9	93.8	0.8	83.1	1.7
Anthracene	96.7	1.0	87.6	0.8	92.1	0.6
Fluoranthene	83.4	1.3	93.9	1.5	95.9	1.2
Pyrene	93.5	1.8	86.1	1.3	95.0	1.4
1,2-Benzanthracene	94.5	1.3	89.6	1.6	94.9	1.0
Chrysene	101.0	1.4	97.8	1.7	87.2	1.6
Benzo[e]pyrene	88.8	1.5	85.2	1.9	95.0	1.4
Benz[e]acenaphthylene	95.5	0.7	92.7	0.7	89.2	0.9
Benzo[k]fluoranthene	93.5	0.8	94.6	0.9	98.9	0.8
Dibenzo[a,h]anthracene	88.2	0.9	97.3	1.1	97.1	0.6
Benzo[g,h,i]perylene	98.4	0.8	95.5	1.6	98.2	0.7
Indeno[1,2,3-cd]pyrene	91.5	1.5	97.9	0.9	94.3	0.7

<sup>\*</sup> UV detection at 230 nm



# **Acrylamides in Fried Foods by QuEChERS Extraction for Acrylamides**

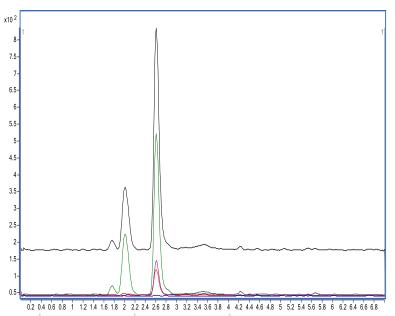




Fadwa Al-Taher, NCFST, IL



# Analysis of Acrylamides in Fried Foods by QuEChERS Extraction for Acrylamides



MRM transitions:		
 TIC		
 12C-AA 72.0 → 55.2		
 12C-AA 72.0 → 26.7		
 13C-AA 75.0 → 58.2		
 13C-AA 75.0 → 29.0		

Matrix	Concentration of acrylamide spike (ng/mL)	%Recovery (n=3)	%RSD (n=3)
1:1 ACN-water	50	116.6	4.07
1:1 ACN-water	100	114.06	4.85
French fries	100	97.14 (after blank correction)	5.04
French fries	200	97.50 (after blank correction)	2.55